SANTA CRUZ BIOTECHNOLOGY, INC.

rabbit serum: sc-2338

BACKGROUND

Santa Cruz Biotechnology offers a wide variety of control immunoglobulin and control sera for a large selection of species, including goat, donkey, rabbit, mouse, rat, bovine, cat, chicken, dog, guinea pig, Syrian hamster, horse, swine, turkey and sheep. Our normal serum contains multiple classes of immunoglobulins and serum proteins from non-immunized animals. Normal serum is provided for use as a blocking reagent to prevent non-specific interactions of tissues or cells in immunohistochemistry, immunocytochemistry and immunofluorescence studies. When used as an antibody diluent in these applications, normal serum provides a ideal, native-like environment. The serum used should be of the same species as that in which the secondary antibody was raised. For example, if using goat anti-rabbit IgG-HRP secondary antibody in a research application, select the normal goat serum as the blocking reagent. Control immunoglobulin and immunoglobulin conjugates are useful negative controls. Santa Cruz Biotechnology offers affinity purified normal immunoglobulins and immunoglobulin conjugates for use as negative controls in applications including flow cytometry, immunohistochemistry and immunofluorescence. Isotype specific control immunoglobulins include classes such as mouse IgG_1,IgG_{2a}, IgG_{2b}, IgG_3, IgM and IgA, rat IgG_1,IgG_{2a}, IgG_{2b} and IgM, Armenian hamster IgG, and both goat and rabbit IgG. Most control immunoglobulins are available as unconjugated controls or as FITC (fluorescein isothiocyanate), PE (phycoerythrin), PE-Cy5 (phycoerythrin-Cy5), PE-Cy7 (phycoerythrin-Cy7), APC (allophycocyanin) and APC-Cy7 (allophycocyanin-Cy7) conjugates. Additional conjugates include Alexa Fluor® 488, Alexa Fluor® 647, Alexa Fluor® 405, PerCP (peridinin chlorophyll protein complex) and PerCP-Cy5.5 (peridinin chlorophyll protein complex-Cy 5.5).

SOURCE

Normal rabbit serum is provided as neat serum from a non-immunized animal.

PRODUCT

Each vial contains 1 ml normal rabbit serum containing < 0.01% Thimerisol.

APPLICATIONS

Normal rabbit serum is recommended for use as a blocking reagent for immunofluorescence, immunohistochemistry and immunocytochemistry. To be used at an assay dependent dilution. In research applications, the species of the normal serum should match the host species of the secondary antibody.

STORAGE

Store at 4° C, **DO NOT FREEZE**. Stable for one year from the date of shipment. Non-hazardous. No MSDS required.

RESEARCH USE

For research use only, not for use in diagnostic procedures.

PROTOCOLS

See our web site at www.scbt.com or our catalog for detailed protocols and support products.

RECOMMENDED SUPPORT PRODUCTS

A. TISSUE CULTURE CELLS

- CrystalCruz™ Cover Glasses, 22 x 50 mm, precleaned: sc-24975
- CrystalCruz™ Micro Slides 75 x 25 mm; 72 frosted sides: sc-24976
- PBS (Phosphate Buffered Saline), powder, 1 packet: sc-24947
- Formaldehyde, 37% formaldehyde solution, 25 ml: sc-203049
- Hydrogen Peroxide, 30% solution, 100 ml: sc-203336

B. FROZEN TISSUE SECTIONS

- Organo/Limonene Mount, non-toxic Permount alternative, 100 ml: sc-45087
- UltraCruz™ Mounting Medium, aqueous-based, 10 ml: sc-24941
- ImmunoHistoMount, aqueous-based mounting medium, 30 ml: sc-45086
- Immuno In Situ Mount, for use with *in situ* hybridization, 30 ml: sc-45088

C. FORMALIN-FIXED, PARAFFIN-EMBEDDED TISSUE SECTIONS

- Paraffin, for the preparation of tissue samples for staining, 500 g: sc-286633
- Xylenes, mixed isomers with ethylbenzene, 500 ml: sc-237422
- Hematoxylin, Gill's Formulation #2; nuclear counter stain, 100 ml: sc-24973

SELECT PRODUCT CITATIONS

1. Demkovich, LE. 1983. 'PPO'—three letters that may form one answer to runaway health costs. Natl. J. 15: 1176-1177.
2. Zhang, L., et al. 1999. Diminished G_1 checkpoint after γ-irradiation and altered cell cycle regulation by Insulin-like growth factor II overexpression. J. Biol. Chem. 274: 13118-13126.
3. Kersh, E.N., et al. 2002. TCR signal transduction in antigen-specific memory CD8 T cells. J. Immunol. 170: 5455-5463.
4. Mattila, P.E., et al. 2005. Cytoskeletal interactions regulate inducible L-selectin clustering. Am. J. Physiol. Cell Physiol. 289: C323-C332.
5. Nozell, S. and Laver, T. 2006. Mechanism of IFN-β-mediated inhibition of IL-8 gene expression in astroglioma cells. J. Immunol. 177: 822-830.
6. Srivastava, K., et al. 2007. 15(S)-hydroxyeicosatetraenoic acid-induced angiogenesis requires STAT3-dependent expression of VEGF. Cancer Res. 67: 4328-4336.
7. Potula, H.S., et al. 2009. Src-dependent STAT-3-mediated expression of monocyte chemoattractant protein-1 is required for 15(S)-hydroxyeicosatetraenoic acid-induced vascular smooth muscle cell migration. J. Biol. Chem. 284: 31142-31155.
8. Singh, NK., et al. 2010. AP-1 (Fra-1/c-Jun)-mediated induction of expression of matrix metalloproteinase-2 is required for 15S-hydroxyeicosatetraenoic acid-induced angiogenesis. J. Biol. Chem. 285: 16830-16843.
9. Liu, J., et al. 2011. G-protein α-s and -12 subunits are involved in androgen-stimulated PI3K activation and androgen receptor transactivation in prostate cancer cells. Prostate. 71: 1276-1286.
10. Thiollier C,, et al. 2012. Characterization of novel genomic alterations and therapeutic approaches using acute megakaryoblastic leukemia xenograft models. J. Exp. Med. 209: 2017-2031.