SANTA CRUZ BIOTECHNOLOGY, INC.

donkey anti-goat IgG-TR: sc-2783

SANTA CRUZ BIOTECHNOLOGY
The Power to Question

BACKGROUND

Santa Cruz Biotechnology's secondary antibodies are available conjugated to either an enzyme, biotin or fluorophore for use in a variety of antibody-based applications including Western Blot, immunostaining, flow cytometry and ELISA. Secondary antibodies are commonly affinity purified against immobilized whole IgG or against antibody fragments. Santa Cruz Biotechnology offers an extensive selection of secondary antibodies optimized for immunohistochemistry and flow cytometry, and are labeled with either biotin, FITC (fluorescein isothiocyanate), Texas Red®, TRITC (tetramethyl rhodamine iso-thiocyanate), PE (phycoerythrin), PerCP (peridinin chlorophyll protein complex) and PerCP-Cy5.5 (peridinin chlorophyll protein complex with cyanin-5.5). Immunohistochemistry and flow cytometry secondary antibodies are specific for commonly used primary antibody species, including goat, rabbit, mouse and rat.

SOURCE

donkey anti-goat IgG-TR is a pre-adsorbed, affinity purified secondary antibody raised in donkey against goat IgG and conjugated to Texas Red®.

PRODUCT

Each vial contains 200 µg donkey IgG (pre-adsorbed with mouse and human IgG) in 0.5 ml of PBS containing 0.02% sodium azide.

APPLICATIONS

donkey anti-goat IgG-TR is recommended for detection of goat IgG by immunofluorescence staining (starting dilution: 1:100, dilution range: 1:100-1:400) and immunohistochemical staining (starting dilution: 1:100, dilution range: 1:100-1:400).

RECOMMENDED SUPPORT PRODUCTS

A. TISSUE CULTURE CELLS

- CrystalCruz™ Cover Glasses, 22 x 50 mm, precleaned: sc-24975
- CrystalCruz™ Micro Slides 75 x 25 mm; 72 frosted sides: sc-24976
- PBS (Phosphate Buffered Saline), powder, 1 packet: sc-24947
- Formaldehyde, 37% formaldehyde solution, 25 ml: sc-203049
- Hydrogen Peroxide, 30% solution, 100 ml: sc-203336

B. FROZEN TISSUE SECTIONS

- Organo/Limonene Mount, non-toxic alternative to Permount, 100 ml: sc-45087
- UltraCruz™ Mounting Medium, aqueous-based, 10 ml: sc-24941
- ImmunoHistoMount, aqueous-based mounting medium, 30 ml: sc-45086
- Immuno *In Situ* Mount, for use with *in situ* hybridization, 30 ml: sc-45088

C. FORMALIN-FIXED, PARAFFIN-EMBEDDED TISSUE SECTIONS

- Paraffin, for the preparation of tissue samples for staining, 500 g: sc-286633
- Xylenes, mixed isomers with ethylbenzene, 500 ml: sc-237422
- Hematoxylin, Gill's Formulation #2; nuclear counter stain, 100 ml: sc-24973

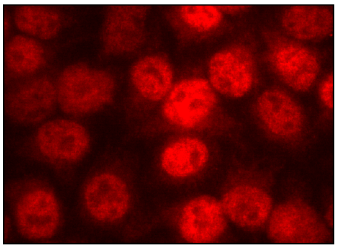
RESEARCH USE

For research use only, not for use in diagnostic procedures.

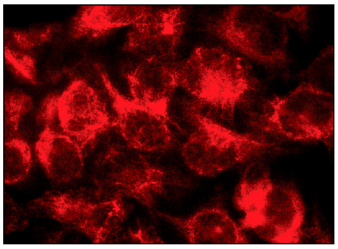
STORAGE

Store at 4° C, **DO NOT FREEZE**. Stable for one year from the date of shipment. Non-hazardous. No MSDS required.

DATA



donkey anti-goat IgG-TR: sc-2783. Immunofluorescence staining of methanol-fixed HeLa cells showing nuclear localization. Antibody tested: Brg-1 (H-88): sc-10768.



donkey anti-goat IgG-TR: sc-2783. Immunofluorescence staining of methanol-fixed HeLa cells showing cytoplasmic localization. Antibody tested: CD89B (G-20): sc-17082.

SELECT PRODUCT CITATIONS

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3. Xiao, Z., et al. 2008. Analysis of the extracellular matrix and secreted vesicle proteomes by mass spectrometry. Methods Mol. Biol. 428: 231-244.
4. Marion, V., et al. 2009. Transient ciliogenesis involving Bardet-Biedl syndrome proteins is a fundamental characteristic of adipogenic differentiation. Proc. Natl. Acad. Sci. USA 106: 1820-1825.
5. Glushakova, L.G., et al. 2009. AAV3-mediated transfer and expression of the pyruvate dehydrogenase E1 α subunit gene causes metabolic remodeling and apoptosis of human liver cancer cells. Mol. Genet. Metab. 98: 289-299.
6. Wilson, C.M., et al. 2009. Radiation-induced astrogliosis and blood-brain barrier damage can be abrogated using anti-TNF treatment. Int. J. Radiat. Oncol. Biol. Phys. 74: 934-941.
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8. Gonzales, A.L., et al. 2010. Ca^{2+} release from the sarcoplasmic reticulum is required for sustained TRPM4 activity in cerebral artery smooth muscle cells. Am. J. Physiol., Cell Physiol. 299: C279-C288.
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10. Chen, Y.F., et al. 2011. Examining rhodopsin retention in endoplasmic reticulum and intracellular localization *in vitro* and *in vivo* by using truncated rhodopsin fragments. J. Cell. Biochem. 112: 520-530.