

normal mouse IgG_{2a}-FITC: sc-2856

BACKGROUND

Santa Cruz Biotechnology offers a wide variety of control immunoglobulin and control sera for a large selection of species, including mouse, rabbit, goat, chicken, rat, hamster, canine, guinea pig and sheep. Control immunoglobulin and immunoglobulin conjugates are useful negative controls. Normal sera is offered to be used as blocking reagents. Santa Cruz Biotechnology offers affinity purified normal immunoglobulins and immunoglobulin conjugates for use as negative controls in applications including flow cytometry, immunohistochemistry, immunofluorescence, Western Blotting and immunoprecipitation. Agarose (AC) conjugated IgGs are provided for immunoprecipitation; horseradish peroxidase (HRP) conjugates are provided for Western Blotting and immunohistochemistry; and Biotin (B) conjugates are provided for immunohistochemistry. A broad range of fluorescent conjugated controls are also available for use in flow cytometry and immunofluorescence applications. Most control immunoglobulins are available as unconjugated controls or as FITC (fluorescein isothiocyanate), PE (phycoerythrin), PE-Cy5 (phycoerythrin-Cy5), PE-Cy7 (phycoerythrin-Cy7), APC (allophycocyanin) and APC-Cy7 (allophycocyanin-Cy7) conjugates. Additional conjugates include Alexa Fluor® 488, Alexa Fluor® 647, Alexa Fluor® 405, PerCP (peridinin chlorophyll protein complex) and PerCP-Cy5.5 (peridinin chlorophyll protein complex-Cy 5.5). Isotype specific control immunoglobulins include classes such as mouse IgG₁, IgG_{2a}, IgG_{2b}, IgG₃, IgM and IgA, rat IgG₁, IgG_{2a}, IgG_{2b} and IgM, Armenian hamster IgG, and both goat and rabbit IgG.

SOURCE

normal mouse IgG_{2a}-FITC is an affinity purified, FITC (fluorescein) conjugated isotype control immunoglobulin from mouse.

PRODUCT

Each vial contains 200 µg mouse IgG_{2a} in 1.0 ml PBS containing 1% stabilizer protein and 0.02% sodium azide.

APPLICATIONS

normal mouse IgG_{2a}-FITC is recommended for use as an isotype control immuno-globulin in place of a target specific primary antibody of the same isotype (mouse IgG_{2a}) by immunofluorescence, immunohistochemical staining (including paraffin-embedded sections) and flow cytometry. To be used at an assay dependent dilution.

STORAGE

Store at 4° C, ****DO NOT FREEZE****. Stable for one year from the date of shipment. Non-hazardous. No MSDS required.

PROTOCOLS

See our web site at www.scbt.com for detailed protocols and support products.

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RECOMMENDED SUPPORT PRODUCTS

- CrystalCruz™ Cover Glasses, 22 x 50 mm: sc-24975
- PBS, powder: sc-24947
- Formaldehyde: sc-203049
- Hydrogen Peroxide: sc-203336
- Organo/Limonene Mount: sc-45087
- UltraCruz® Mounting Medium: sc-24941
- ImmunoHistoMount: sc-45086
- Immuno In Situ Mount: sc-45088
- Paraffin: sc-286633
- Xylenes: sc-237422
- Hematoxylin: sc-24973
- FCM Lysing solution: sc-3621
- FCM Fixation Buffer: sc-3622
- FCM Permeabilization Buffer: sc-3623
- FCM Wash Buffer: sc-3624
- Intracellular FCM System: sc-45063

SELECT PRODUCT CITATIONS

1. Ben-Haroush, A., et al. 2005. Expression of basic fibroblast growth factor and its receptors in human ovarian follicles from adults and fetuses. *Fertil. Steril.* 84 Suppl. 2: 1257-1268.
2. Nagai, K., et al. 2009. The hydroxyflavone, fisetin, suppresses mast cell activation induced by interaction with activated T cell membranes. *Br. J. Pharmacol.* 158: 907-919.
3. Murakami, M., et al. 2013. The use of granulocyte-colony stimulating factor induced mobilization for isolation of dental pulp stem cells with high regenerative potential. *Biomaterials* 34: 9036-9047.
4. Yuan, F., et al. 2014. Human liver cell trafficking mutants: characterization and whole exome sequencing. *PLoS ONE.* 9: e87043.
5. Murakami, M., et al. 2015. Trophic effects and regenerative potential of mobilized mesenchymal stem cells from bone marrow and adipose tissue as alternative cell sources for pulp/dentin regeneration. *Cell Transplant.* 24: 1753-1765.
6. Nguyen, M.L., et al. 2016. Dynamic regulation of permissive histone modifications and GATA3 binding underpin acquisition of granzyme A expression by virus-specific CD8⁺ T cells. *Eur. J. Immunol.* 46: 307-318.
7. Hirose, Y., et al. 2016. Injection of dental pulp stem cells promotes healing of damaged bladder tissue in a rat model of chemically induced cystitis. *Cell Transplant.* 25: 425-436.
8. Hirose, Y., et al. 2016. Effects of extracellular pH on dental pulp cells *in vitro*. *J Endod.* 42: 735-741.

RESEARCH USE

For research use only, not for use in diagnostic procedures.